ESQUITERPENE LACTONES, FLAVONOIDS AND COUMARINS FROM CENTAUREA COLLINA

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(Received 5 January 1989)

Key Word Index—Centaurea collina subsp. collina; Compositae; sesquiterpene lactones; guaianolides; flavonoids; coumarins; ¹³C NMR.

Abstract—The isolation and characterization of two coumarins, five flavonoids and four sesquiterpene lactones from the aerial parts of *Centaurea collina* are reported. The ¹³C NMR data are included.

INTRODUCTION

Centaurea collina L. subsp. collina is a herbaceous plant with pale yellow flowers which occurs in the Iberian Peninsula, the south of France and Italy [1]. It belongs to an extensively studied genus, from which many metabolites, mainly sesquiterpene lactones [2] and flavonoids [3], have been isolated. From C. collina we have already reported two guaianolides [4] and Voirin et al. have reported several flavonoids [5-7]. We now report the isolation of further sesquiterpene lactones and several flavonoids and coumarins and aromatic acids. We also report the ¹H-¹³C heteronuclear correlation for two guaianolides.

RESULTS AND DISCUSSION

Chromatographic separation of the ether-soluble portion of a methanolic extract afforded, in addition to benzoic, vanillic and protocatechuic acids, two coumarins identified as scoparone (1) and scopoletin (2) by spectroscopic methods [8], including ¹³C NMR [9, 10], five flavonoids and four sesquiterpene lactones.

The flavonoids isolated were identified [11, 12] as pectolinarigenin (3), hispidulin (4), apigenin (5), eupafolin (6) and luteolin (7). The position of the methoxyl group at C-6 in the three 6-methoxyflavones 3, 4 and 6 followed from the UV [13] and the mass spectral [14] data. As far as we know, the ¹³C NMR spectra of compounds 3 and 6 have not been described in the literature so far. Although the ¹³C NMR spectrum of compound 4 has already been reported [3], it is also included for comparison (Table 1). The assignment of the signals was based on the work of Mabry et al. [15] and distortionless enhancement by polarization transfer (DEPT) experiments. A CH signal at δ 94 was assignable to the unsubstituted C-8; this value is in the characteristic zone (δ 90–95) of this carbon when it is not substituted [3, 16]. A quaternary signal at δ 131 was assigned to C-6 substituted by a methoxyl group. It is interesting to note that the flavonoids isolated by us are flavones, while the flavonoids isolated by Voirin et al. [5-7] are flavonols, except for pectolinarigenin.

The four sesquiterpene lactones isolated belong to the guaianolides group. The structures of two of them (8 and 9), we have reported previously [4] as 3β -hydroxy-8 α epoxymethyl acriloiloxy-4(15),10(14),11(13)-trien- $(1\alpha H)$, $(5\alpha H)$ -guaian-6,12-olide and its 11β ,13-dihydroderivative respectively. In that report [4], we characterized the two guaianolides by spectroscopic methods, including DEPT techniques for ¹³C NMR. Nevertheless, some resonances could not be assigned without ambiguity. We have now resolved these ambiguities by ¹H-¹³C heteronuclear correlation (Table 2). Thus, in compound 8, the signal at δ 78.1 is assigned to C-6 by its correlation with the triplet at δ 4.22 characteristic of H-6 and the signal at δ 75.2 is assigned to C-8 as it is correlated with the doublet triplet at δ 5.05 of H-8. In the same way, the signals at δ 45.3 and 46.9 are assigned at C-1 and C-7 by their correlation with the quadruplet at δ 2.91 for H-1 and the triplet triplet at δ 3.06 of H-7, respectively. Similarly the signals at δ 38.3 and 35.6 are assigned at C-2 and C-9 as they are correlated with the signals at $\delta 1.68$ and 2.11 of H-2 and 2.30 and 2.60 of H-9, respectively. In compound 9, the ambiguities in the ¹³C NMR spectrum interpretation were resolved by ¹H-¹³C heteronuclear correlation also. Thus, the signals at δ 51.0 and 52.2 are assigned at C-5 and C-7 respectively by their correlation with the triplet at $\delta 2.77$ of H-5 and the quadruplet at $\delta 2.24$ of H-7. The other ambiguities of the 13C NMR spectrum interpretation, were similarly resolved.

The remaining two guaianolides were identified as deacylcynaropicrin (10) [17] and its 11β ,13-dihydroderivative 11 [18] by spectral data as well as by comparison with the corresponding published data [19]. We include the ¹³C NMR data (Table 2), as it has not been described in the literature so far. Of the guaianolides isolated, the deacylcynaropicrin 10 is widely distributed in Compositae, whilst its 11β ,13-dihydroderivative (11) has been reported previously from Tricholepis glaberrima [18], Saussurea affinis [20] and Centaurea canariensis [21]. Compound 8 has been isolated previously from Centaurea solstitialis [22]. Its 11β ,13-dihydroderivative (9) has been reported by us for the first time from C. collina [4].

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3
$$R^1 = R^2 = OMe, R^3 = H$$

4 $R^1 = OMe, R^2 = OH, R^3 = H$
5 $R^1 = R^3 = H, R^2 = OH$
6 $R^1 = OMe, R^2 = R^3 = OH$

$$8 \quad R = \frac{18}{16} O$$

$$10 R = H$$

9 R =
$$\frac{18}{17}$$

11
$$R = H$$

Table 1. ¹³C NMR of flavonoids 3, 4 and 6 from C. collina

C	3	4	6
2	163.34	163.76	164.10
3	103.04	102.36	102.54
4	182.14	182.10	182.16
5	152.75a	152.75°	152.94
6	131.37	131.31	131.48
7	157.33	157.25	157.35
8	94.30	94.21	94.24
9	152.41ª	152.36°	152.50
10	104.12	104.03	104.22
1'	122.85	121.19	119.08
2'	128.26	128.43	113.49
3'	114.56	115.92	145.87
4'	162.29	161.13	149.81
5'	114.56	115.92	116.15
6'	128.26	128.43	121.70
-OMe (C ₆)	59.93	59.92	60.07
-OMe (C _{6'})	55.54		

^aChemical shifts in each column may be interchanged.

Table 2. ¹³C NMR of guaianolides **8–11** from *C. collina*

C	8	9	10	11
1	45.3	44.5	45.2ª	44.2°
2	38.3	38.7	39.2ь	39.0 ^t
3	73.2	73.3	73.7	73.6
4	151.8	152.4	152.4	153.0
5	51.5	51.0	51.3ª	50.7ª
6	78.1	78.7	79.0	79.1
7	46.9	52.2	51.0ª	56.0°
8	75.2	77.7	71.9	74.9
9	35.6	39.0	41.3 ^b	44.8 ^t
10	141.6	141.8	142.7	143.2
11	136.8	41.3	138.1	42.0
12	169.5	178.4	169.9	
13	123.6	15.2	123.2	15.9
14	118.1	117.5	117.1	116.2
15	113.8	112.8	113.2	112.0
16	174.8	175.0	mandon.	
17	76.0	75.9		
18	68.1	68.0	about The	
19	21.6	21.8		

a.b. Chemical shifts denoted by the same letter in each column may be interchanged.

EXPERIMENTAL

Centaurea collina was collected, classified and extracted as described previously [4]. The Et₂O re-extract was subjected to chromatography on a silica gel column using mixtures of hexane-CH₂Cl₂-EtOAc as eluent. Six main groups (I-VI) of fractions were obtained, with the following proportions of CH₂Cl₂-EtOAc: I (19:1), II (19:1 to 9:1), III (17:3), IV (4:1 to 7:3), V (7:3 to 1:1) and VI (9:11). Compound 2 (65 mg) and 4 (59 mg) crystallized directly from the group II and IV fractions respectively. Repeated chromatographic process afforded: compound 1 (12 mg) from group I; benzoic acid (23 mg) and compound 3 (22 mg) from group III; vanillic acid (71 mg) and compound 5 (19 mg) from the mother liquors of group IV; protocatechuic acid (26 mg) and compounds 10 (10 mg), 11 (5 mg), 6 (14 mg) and 7 (10 mg) from group V and compounds 8 (30 mg) and 9 (15 mg) from group VI.

Acknowledgements—Financial support from the Comisión Asesora de Investigación Cientifica y Tècnica (CAICYT, Grant No. 559/84) is gratefully acknowledged. We thank Prof. Dr J. Mansanet (Department of Botany, Faculty of Biological Sciences, University of Valencia, Spain) for identification of plant material.

REFERENCES

- Flora Europaea (1976) (Tutin, T. G., Heywood, V. H., Burger, N. A., Moore, D. M., Valentine, D. H., Walters, S. M. and Webb, D. A. eds) Vol. 4, p. 263. Cambridge University Press, Cambridge.
- 2. Fraga, B. M. (1987) Nat. Prod. Reports 4, 473.
- 3. Gonzalez Collado, I., Macias, F. A., Massanet, G. M. and Rodriguez Luis, F. (1985) J. Nat. Prod. 48, 819.
- Fernández, I., García, B., Grancha, F. J. and Pedro, J. R. (1987) Phytochemistry 26, 1403.

- Kamanzi, K., Raynaud, J. and Voirin, B. (1982) Pharmazie 37, 454.
- Kamanzi, K., Raynaud, J. and Voirin, B. (1982) Plant. Med. Phytother. 16, 30.
- Kamanzi, K., Raynaud, J. and Voirin, B. (1982) Pharmazie 37, 523.
- Barberá, O., Marco, J. A., Sanz, J. F. and Sánchez-Parareda, J. (1986) Phytochemistry 25, 2357.
- Sankar, S. S., Gilbert, R. D. and Fornes, R. E. (1982) Org. Magn. Reson. 19, 223.
- Günther, H., Prestien, J. and Nathan, P. J. (1975) Org. Magn. Reson. 7, 339.
- 11. Mabry, T. J., Markham, K. R. and Thomas, M. B. (1970) The Systematic Identification of Flavonoids. Springer, New York.
- 12. Voirin, B. (1983) Phytochemistry 22, 2107.
- 13. Mears, J. A. and Mabry, T. J. (1972) Phytochemistry 11, 411.
- 14. Goudard, M., Favre-Bonvin, J., Lebreton, P. and Chopin, J. (1978) Phytochemistry 17, 145.
- Markham, K. R., Chari, V. M. and Mabry, T. J. (1982) The Flavonoids: Advances in Research (Harborne, J. B. and Mabry, T. J., eds) Chap 2. Chapman & Hall, London.
- Chari, V. M., Ahmad, S. and Osterdahl, B. G. (1978) Z. Naturforsch. 33b, 1547.
- González, A. G., Bermejo, J., Massanet, G. M. and Perez, J. (1973) An. Quim. 69, 1333.
- Singhal, A. K., Chowdhury, P. K., Sharma, R. P., Baruah, J. N. and Herz, W. (1982) Phytochemistry 29, 462.
- González, A. G., Bermejo, J., Castañeda, J. and Estévez, F. (1985) Phytochemistry 24, 1383.
- Das, S., Baruah, R. N., Sharma, R. P., Baruah, J. N., Kulanthaivel, P. and Herz, W. (1983) Phytochemistry 22, 1989
- González, I., Macias, F. A., Massanet, G. M. and Rodriguez, F. (1985) Phytochemistry 24, 2107.
- Jakupovic, J., Jia, Y., Pathak, V. P., Bohlmann, F. and King, R. M. (1986) Planta Med. 399.